## Insecticidal 4-Alkylidene-1,3,2-benzodioxaphosphorinane Derivatives


#### Abstract

2-Alkoxy-4-alkylidene-1,3,2-benzodioxaphosphorin 2-sulfides and related compounds are obtained from $o$-hydroxyacetophenone and other o-alkylketophenols by several routes, usually involving thermal cyclization of the appropriate phosphorochloridate intermediates. ${ }^{1} \mathrm{H}$ and ${ }^{31} \mathrm{P}$ NMR and HPLC studies assign the higher 4 -alkylidene derivatives as the $Z$-form configuration. As in the salithion series, cyclic phosphorus esters with lower 2-alkoxy or 2-alkylamino and 4-alkylidene substituents are potent insecticides.


The potency of salithion (1) (Eto et al., 1963) and related


saligenin cyclic phosphorus esters (Eto, 1974) as insecticides prompted synthesis and bioassay of analogous compounds ( $2, R_{1}, R_{2}$, and $R_{3}=$ lower alkyl) with 4 -alkylidene substituents.

Preparation of Cyclic Phosphorus Esters. Synthesis routes are given in Figure 1 with examples of the products in Table I. Compound $3\left(\mathrm{R}_{1}=\mathrm{H}, \mathrm{CH}_{3}\right)$ from commercial sources was converted to $4\left(\mathrm{R}_{1}=\mathrm{H}, \mathrm{CH}_{3}\right)$ and $5\left(\mathrm{R}_{1}=\mathrm{H}\right.$, $\mathrm{CH}_{3} ; \mathrm{R}_{2}=\mathrm{CH}_{3}, \mathrm{C}_{2} \mathrm{H}_{5}$ ) by a reported procedure (Cragg et al., 1977). Cyclic phosphates 6 and 7 were obtained by reaction of $5\left(\mathrm{R}_{1}=\mathrm{H} ; \mathrm{R}_{2}=\mathrm{CH}_{3}, \mathrm{C}_{2} \mathrm{H}_{5}\right)$ with equimolar $m$-chloroperbenzoic acid (MCPBA) in dichloromethane at $-50^{\circ} \mathrm{C}$ with product isolation by preparative TLC (silica gel; hexane-acetone, 7:2). Cyclic phosphorothionates 9-11 were prepared by stirring a mixture of 5 and equimolar elemental sulfur in carbon disulfide for 5 h at $50^{\circ} \mathrm{C}$ and isolating the products by distillation. Two other routes also provided the cyclic phosphorothionates. Compounds 12 and 14 were obtained by mixing equimolar $3\left(\mathrm{R}_{1}=\mathrm{CH}_{3}\right.$, $\mathrm{C}_{2} \mathrm{H}_{5}$ ) with O-ethyl thionophosphoryl dichloridate in $20 \%$ NaOH at $10^{\circ} \mathrm{C}$, followed by stirring for 2 h at $25^{\circ} \mathrm{C}$ and recovery by extraction into chloroform. Slow distillation under reduced pressure converted the intermediate 8 ( $\mathrm{R}_{1}$ $=\mathrm{CH}_{3}, \mathrm{C}_{2} \mathrm{H}_{5} ; \mathrm{R}_{2}=\mathrm{C}_{2} \mathrm{H}_{5}$ ) to 12 and 14. The SchottenBaumann procedure as above but with thiophosphoryl trichloride gave $15\left(\mathrm{R}_{1}=\mathrm{H}, \mathrm{CH}_{3}\right)$ which was converted to $16\left(\mathrm{R}_{1}=\mathrm{H}, \mathrm{CH}_{3}\right)$ on distillation ( $\sim 135^{\circ} \mathrm{C} / 0.1 \mathrm{mmHg}$ ). This was a very useful intermediate, providing 13 on reaction with equimolar propanol and triethylamine in benzene and giving 17 and 18 by addition of excess amine to the chloridate in benzene at $25^{\circ} \mathrm{C}$. Compounds 13,17 , and 18 were isolated by preparative TLC as above. The


Figure 1
reported yields refer to the final step in the reaction sequence.

Purity and Structure. All compounds (including 4, $5,8,15$, and 16) appeared to be pure by TLC and gave appropriate chemical ionization quasi-molecular ions (M $+1)^{+}$and the anticipated ${ }^{1} \mathrm{H}$ NMR spectra in $\mathrm{CDCl}_{3}$ (tetramethylsilane). ${ }^{31} \mathrm{P}$ NMR spectra were obtained for solutions in $\mathrm{CDCl}_{3}$, reporting chemical shifts as positive when downfield of $1 \%$ trimethyl phosphate in $\mathrm{CDCl}_{3}\left({ }^{31} \mathrm{P}\right.$ $=0$ ). The higher alkylidene derivatives 11-14 each gave a single spot on TLC and a single peak on HPLC ( $\mu$ Porasil; hexane-chloroform, 4:1). ${ }^{1} \mathrm{H}$ NMR and ${ }^{31} \mathrm{P}$ NMR (16, $\mathrm{R}_{1}$ $=\mathrm{CH}_{3} ;{ }^{31} \mathrm{P}=46.5 \mathrm{ppm}$; see also Table I) further supported the presence of a single isomer. ${ }^{1} \mathrm{H}$ NMR assigned the $Z$-form configuration. The signals for $\mathrm{H}_{\mathrm{a}}(5.64-5.76 \mathrm{ppm})$ appeared as a double quartet ( $\mathrm{R}_{1}=\mathrm{CH}_{3} ; J_{\mathrm{H}_{-}-\mathrm{CH}}=7 \mathrm{~Hz}$ ) or a double triplet ( $\mathrm{R}_{1}=\mathrm{C}_{2} \mathrm{H}_{5} ; J_{\mathrm{H}_{-}-\mathrm{CH}}=7 \mathrm{~Hz}$ ) because of the coupling with phosphorus ( ${ }^{4} J_{\mathrm{P}-\mathrm{H}_{\mathrm{A}}}=5 \mathrm{~Hz}$ for phosphates and 3 Hz for phosphorothionates) [ $W$ effect (Cooper, 1980)]. It seems likely that a high degree of stereoselectivity is involved in the enolization and cyclization reaction sequence.
Biological Activity. The toxicity of the new compounds applied topically to adult female houseflies (Musca

Table I. Properties of 4-Alkylidene-1,3,2-benzodioxaphosphorinane Derivatives

| no. | phosphorus moiety of 2 | $\mathrm{H}_{\mathrm{b}}$ or $\mathrm{R}_{1}$ | yield, \% | $\mathrm{mp},{ }^{\circ} \mathrm{C}$, or $\mathrm{bp},{ }^{\circ} \mathrm{C} / \mathrm{mmHg}$ | ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right), \delta$ |  | $\begin{gathered} { }^{31} \mathrm{P} \text { NMR } \\ \left(\mathrm{CDCl}_{3}\right), \delta \end{gathered}$ | $\begin{gathered} \text { fly } \operatorname{LD}_{50}, \\ \mu \mathrm{~g} / \mathrm{g} \end{gathered}$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  |  |  | $\mathrm{H}_{\mathrm{a}}$ | $\mathrm{H}_{\mathrm{b}}$ |  |  |
| 6 | $\mathrm{P}(\mathrm{O}) \mathrm{OCH}_{3}$ | H | 80 | 27-28 | 5.23 | 5.00 | -15.7 | 8 |
| 7 | $\mathrm{P}(\mathrm{O}) \mathrm{OC}_{2} \mathrm{H}_{5}$ | H | 85 | 25-27 | 5.29 | 5.04 | -16.5 | 43 |
| 9 | $\mathrm{P}(\mathrm{S}) \mathrm{OCH}_{3}$ | H | 90 | 125-130/0.1 | 5.24 | 5.00 | 52.0 | 7 |
| 10 | $\mathrm{P}(\mathrm{S}) \mathrm{OC}_{2} \mathrm{H}_{5}$ | H | 92 | 134-136/0.1 | 5.21 | 4.94 | 51.0 | 18 |
| 11 | $\mathrm{P}(\mathrm{S}) \mathrm{OCH}_{3}$ | $\mathrm{CH}_{3}$ | 85 | 135-137/0.1 | 5.76 |  | 53.6 | 56 |
| 12 | $\mathrm{P}(\mathrm{S}) \mathrm{OC}_{2} \mathrm{H}_{5}$ | $\mathrm{CH}_{3}$ | 20 | 140-142/0.1 | 5.70 |  | 51.9 | 43 |
| 13 | $\mathrm{P}(\mathrm{S}) \mathrm{OC}_{3} \mathrm{H}$, | $\mathrm{CH}_{3}$ | 20 | oil | 5.69 |  | 51.9 | 590 |
| 14 | $\mathrm{P}(\mathrm{S}) \mathrm{OC}_{2} \mathrm{H}_{5}$ | $\mathrm{C}_{2} \mathrm{H}_{5}$ | 35 | 140-142/0.1 | 5.64 |  | 52.0 | 15 |
| 17 | $\mathrm{P}(\mathrm{S}) \mathrm{NHCH}_{3}$ | H | 90 | oil | 5.22 | 4.94 | 58.2 | 7 7 |
| 18 | $\mathrm{P}(\mathrm{S}) \mathrm{NHC}_{3} \mathrm{H}_{7}$ | H | 19 | oil | 5.24 | 4.89 | 61.8 | 240 |

domestica L., SCR strain) was somewhat less than that of salithion and its ethoxy analogue which gave $\mathrm{LD}_{50}$ values of 2.5 and $7 \mu \mathrm{~g} / \mathrm{g}$, respectively. Within the 4 -alkylidene series considered here, high toxicity to houseflies was conferred by a phosphate or phosphorothionate moiety, by several small exocyclic substituents (e.g., methoxy, ethoxy, or methylamino), and by $\mathrm{R}_{1}$ as $\mathrm{H}, \mathrm{CH}_{3}$, or $\mathrm{C}_{2} \mathrm{H}_{5}$. The potency of compound 6 in inhibiting fly head and electric eel acetylcholinesterase was similar to that of the phosphate analogue of salithion. Several $O$-phenyl cyclic phosphates ( $\mathrm{R}_{1}=\mathrm{H}, \mathrm{CH}_{3}$, or $\mathrm{C}_{2} \mathrm{H}_{5} ; \mathrm{R}_{2}=$ phenyl) administered intraperitoneally were potent synergists for malathion toxicity in mice and were delayed neurotoxicants in hens.

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## CORRESPONDENCE

## On Globulins of Soybean Seeds

Sir: My colleagues and I at Iowa State University wish to report some serious deletions in the literature citations in a pair of articles appearing in the Journal of Agricultural and Food Chemistry. The articles are the following: " 2 S Globulins of Soybean Seeds. 1. Isolation and Characterization of Protein Components", by I. Koshiyama,* M. Kikuchi, K. Harada, and D. Fukushima, J. Agric. Food Chem. 1981, 29, 336; "2S Globulins of Soybean Seeds. 2. Physicochemical and Biological Properties of Protease Inhibitors", by I. Koshiyama, M. Kikuchi, and D. Fukushima, J. Agric. Food Chem. 1981, 29, 340. Nowhere in their literature citations do they cite the original work of others more than 15 years ago. Among the original works are the following.

1. Birk, Y., et al. Biochem. J. 1963, 87, 281-284.
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Shinkichi Tawata
Masayoshi Eto ${ }^{1}$
John E. Casida*

Pesticide Chemistry and Toxicology Laboratory
Department of Entomological Sciences
University of California
Berkeley, California 94720
${ }^{1}$ Present address: Laboratory of Food Chemistry Himeji Women's College
1-1-12 Shinzaike-Honcho, Himeji, Hyogo 670, Japan

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10. Rackis, J. J., et al. Arch. Biochem. Biophys. 1962, 98, 471-478.
11. Rackis, J. J., et al. Biochem. Biophys. Res. Commun. 1964, 15, 230-235.
12. Rackis, J. J., et al. Arch. Biochem. Biophys. 1970, 98, 471-478.
There have also been a number of excellent reviews on the chemical and physical properties of these soy trypsin inhibitors. Among them are the following.

Birk, Y. Ann. N.Y. Acad. Sci. 1968, 148, 388-399.
Vogel, R., et al. "Natural Proteinase Inhibitors"; Academic Press: New York, 1968.
Liener, I. E.; Kakade, M. L. In "Toxic Constituents of Plant Foodstuffs"; Liener, I. E., Ed.; Academic Press: New York, 1969.
Kassell, B. Methods Enzymol. 1970, 19, 839-890. The failure to cite at least some of these original workers in the newly published articles leads one to believe the work is "new". Careful examination of the literature shows this to be quite the opposite.

## P. A. Murphy

Department of Food Technology
Iowa State University
Ames, Iowa 50011

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